



# Plagiarism Checker X Originality Report

**Similarity Found: 31%**

Date: Jumat, Februari 01, 2019

Statistics: 924 words Plagiarized / 2941 Total words

Remarks: Medium Plagiarism Detected - Your Document needs Selective Improvement.

---

\_ Survival and growth of stiped snakehead fish (*Channa striata* Bloch.) juvenile reared in acid sulfata water and rainwater medium 1,2Purnamawati, 3Daniel Djokosetiyanto, 3Kukuh Nirmala, 3Enang Harris, 4Ridwan Affandi 1 Study Program of Aquaculture, Graduate School, Bogor Agricultural University, Bogor, West Java, Indonesia; 2 Study Program of Aquaculture, Department of Marine Science and Fisheries, Pontianak State Polytechnic, Jalan Ahmad Yani, West Borneo, Indonesia; 3 Department of Aquaculture, Faculty of Fisheries and Marine Science, Bogor Agricultural University, Bogor, West Java, Indonesia; 4 Department of Aquatic Resources Management, Faculty of Fisheries and Marine Science, Bogor Agricultural University, Bogor, West Java, Indonesia. Corresponding author: Purnamawati, purnamawati\_za@yahoo.com  
Abstract.

The aim of this study was to evaluate the effects of the use of the different mediums on the growth and survival of the striped snakehead fish (*Channa striata*). This experiment applied a completely randomized design with acid sulfata water medium and rainwater medium as the experimental treatments, each treatment had twelve replications. The *C. striata* juvenile with an average initial length of  $2.4 \pm 0.2$  cm and an average initial weight of  $0.21 \pm 0.05$  g reared in the aquariums sizing 30 x 25 x 35 cm (with a water volume of 25 L) with a stocking density of 2 individuals/L, for 40 days.

The specimens were fed commercial feed, twice a day (morning and afternoon) until to apparent satiation. The water replacement was performed every 2 days as much as 10% water of the total water volume in the aquarium. The results showed that rainwater significantly affected the biometric and physiological responses of the *C. striata* juvenile.

The rainwater medium gave better results shown by the higher value of survival (73.89%), growth rate (4.40% day<sup>-1</sup>), feed efficiency (59.1%), protein retention (24.31%), energy retention (41.34%), blood albumin content (3.75 g/100 mL) and blood glucose (26.45 mg/100 mL). Key Words: water quality, air-breathing fish, marginal lands, tidal area. Introduction.

Aquaculture sector in Indonesia is growing up with advances in other sectors that support the continuity of aquaculture practices. The development of aquaculture sector is not only conducted in freshwater area, but also in brackishwater or marine areas. Most of aquaculture practices have been applied in marginal lands, but in general the productivity has still relatively been low.

The low aquaculture productivity in these areas is due to various problems, especially poor water quality. The results of the laboratory analysis in 2014 showed that the water in the tidal area had a pH range of 2.53-3.39, sulfate range at 6.91-8.7 mg L<sup>-1</sup>, Fe range at 0.72- 2.83 mg L<sup>-1</sup>, dissolved oxygen level <5 mg L<sup>-1</sup>, and a large salinity shock, that can be problems for the stenohalin fish (Purnamawati 2017).

The entrance of seawater causes large salinity range in tidal area between the rainy season and the dry season reaching 0- 28 ppt. With this suboptimal medium, not all fisheries commodities can be cultured in the tidal water, especially in the water with acid sulfate soil. Acid sulfate soils are derived from marine and estuarine sediments which show a severe acidification because of the oxidation of sulfides that leads to the formation of sulfuric acid. The water with acid sulfate soil can be recognized by the very low pH values that are less than 4 in water.

This condition leads to several problems, including the insufficient growth of algae, the poor condition and slow growth of the fish, and the hazard of sudden fish mortality during heavy rains after a long dry period (Singh & Poernomo 1984). To determine fisheries commodities which can survive in acid sulfate water, it can be seen from several aspects, including the tolerance to the environment, survival and growth (El-Sayed et al 1996).

The striped snakehead fish (*Channa striata*) is an Indonesian freshwater native species which is widely distributed in Sumatra, Kalimantan and Java Islands. This species has wide range habitats found in rivers, swamps, ponds, canals, lakes, and land of rice fields (Song et al 2013). The local and international markets greatly demand on this fish due to its rapid growth, tolerance to high stocking density, and medicinal value in enhancing wound healing and reducing postoperative pain (Jais et al 1997; Qin et al 1997; Mollah et al 2009).

This fish also has a high tolerance to adverse environments due to its hardness to survive in water with high ammonia content and a low dissolved oxygen level caused by its air-breathing capability assisted by a suprabranchial chamber (Ng & Lim 1990; Qin et al 1997; Chandra & Banerjee 2004; Mollah et al 2009). The growth of the fish is strongly influenced by environmental factors, including nutrients in the rearing medium and water quality parameters such as the optimum salinity, pH, and oxygen level. To get the proper medium for the fish life, it would require a study about the response of the *C. striata* juvenile reared in the different mediums, e.g. acid sulfate water and rainwater. This is because each medium has different nutrient compositions between one medium to other mediums for the cultured fish. The aim of this study was to analyze the effects of different rearing mediums on growth and survival of *C. striata*. Material and Method.

This study was conducted on June-September 2014. The rearing of specimens (*C. striata* juveniles) was conducted at Wet Laboratory, Study Program of Aquaculture, Department of Marine Science and Fisheries, Pontianak State Polytechnic, Pontianak, Indonesia.

The measurement of blood albumin levels, the proximate analysis of the fish feed and the proximate analysis of the experimental fish were conducted at Fish Nutrition Laboratory, Department of Aquaculture, Faculty of Fisheries and Marine Science, Bogor Agricultural University, Bogor, Indonesia. The measurement of plasma glucose levels was conducted at Environmental and Food Technology Laboratory, Tanjungpura University, Pontianak, Indonesia.

Experimental design. This study was conducted in laboratory through a completely randomized design (CRD). The treatments applied in this study consisted of two treatments, acid sulfate water medium and rainwater medium. Each treatment applied 12 replicates. Experimental fish. The experimental fish used in this study were *C. striata* juveniles with an average initial length of  $2.4 \pm 0.2$  cm and an average initial weight of  $0.21 \pm 0.05$  g. The fish were reared in mediums equipped with aeration equipment. Experimental tanks and mediums. The tanks used were 24 units of glass aquarium sizing 30 x 25 x 35 cm. The aquariums were filled with acid sulfate water obtained from tidal land of Kubu Raya District West Borneo and rainwater which had been prepared in a fiber tank.

The tops of aquariums were covered with nets to avoid the fish jumping out of the aquariums. Experimental procedures. Adaptation or acclimatization was performed prior that the experimental fish to be used for the experiment. The objective of this step was to adapt or acclimatize the fish to the new environmental conditions. This step was performed in 4 units of aquarium sizing 30 x 25 x 35 cm for 7 days. *C. striata* which had been acclimatized, were reared in the aquariums with a stocking density of

2 individuals/L (Vivekanandan 1977; Hidayatullah et al 2015). The study was conducted for 40 days.

The fish in all treatment groups were fed with commercial feed with chemical composition based on the result of the proximate analysis presented in Table 1, twice a day until to apparent satiation. Every two days, a water replacement was performed by replacing 10% water of the total water volume in the medium with new water.

Table 1 The result of the proximate analysis of commercial feed used in the present study  
Chemical composition \_Content (% dry weight) \_Protein \_34.86 \_Fat \_3.92 \_Carbohydrate \_42.49 \_Nitrogen free extract \_39.30 \_Crude fiber \_3.19 \_Ash \_8.60 \_Moisture \_11.08 \_  
The fish mortality observation was conducted every day to get the survival data of the experimental fish at the end of the study, while measurements of the length and the weight of the fish were done every 10 days to get the growth data. To record the feed intake was done by summing the daily feed consumed during the study.

Observations and measurements of temperature, pH and dissolved oxygen level were carried out every day, while the measurements of sulfate (SO<sub>4</sub><sup>2-</sup>) and ammonia were performed at the beginning and the end of the study. Experimental parameters. The measurement of water quality parameters were performed following the procedures described by APHA (1989).

The measurement of blood albumin levels followed the method described by Infusino & Panteghini (2013), while the proximate analysis of the fish feed and the experimental fish were carried out according to the procedure by Takeuchi (1988). The measurement of plasma glucose levels was performed using a liquicolor Glucose commercial kit GOD-PAP with calorimetric method and the results of the measurement were read with a spectrophotometer at a wavelength of 500 nm following the procedure by Wedemeyer & Yasutake (1977). Survival rate was calculated using a formula according to Kang'ombe & Brown (2008):  $SR = (N_t \times N_0 - 1) \times 100$  Where: SR - survival (%); N<sub>t</sub> - the number of the fish at the end of the study (individuals); N<sub>0</sub> - the number of the fish at the beginning of the study (individuals). Specific growth rate, was calculated using a formula stated by Weatherley & Gill (1989).  $SGR = [(\ln W_2 - \ln W_1) / (t_2 - t_1)] \times 100$  Where: SGR - specific growth rate (% day<sup>-1</sup>); W<sub>1</sub> - the average weight of the fish at the beginning of the study (g); W<sub>2</sub> - the average weight of the fish at time t<sub>2</sub> (g); t<sub>2</sub> - t<sub>1</sub> - experimental duration. The feed efficiency of the *C. striata* juvenile was calculated using a formula constructed by Kang'ombe & Brown (2008).

$$e = [(W_t + D) - W_o] / F \times 100$$

Where: e - feed efficiency;  $W_0$  - the weight of the fish at the beginning of the study (g);  $W_t$  - the weight of the fish at the end of the study (g); D - the weight of dead fish during the study (g); F - the weight of total feed given during the study (g dry weight). Statistical analysis. The data of survival, specific growth rate, blood albumin level, feed efficiency, protein retention, energy retention, and blood glucose level were analyzed through t test to see the differences among treatments. Results and Discussion.

The data obtained from the measurement of physical-chemical parameters of the water are presented in Table 2, while the data obtained from the measurement of biometric parameters (survival, growth, blood albumin level and feed efficiency) and physiological parameter (blood glucose level) of the *C. striata* juvenile reared in acid sulfate water and rainwater mediums are presented in Table 2.

Based on the data presented in Table 2, it was indicated that the values of water quality parameters of acid sulfate water medium used as a treatment for the *C. striata* juvenile were not appropriate for the species physiological requirements. From all parameters observed, only ammonia, oxygen level and temperature of the water were in the optimum range, while the other parameters were not in the optimum range. However, these average ranges of physical-chemical parameters were still quite good for the survival of *C. striata* juveniles, because these ranges were still on the tolerance ranges. On the other hand, the values of water quality parameters of the rainwater medium used as a treatment for *C. striata* specimens within the optimum ranges. Table 2 The values of physical-chemical parameters of the water in each treatment during the study

Parameters	Treatments	Acid sulfate water	Rainwater	Tolerance range*	optimum range**
pH		3.7–5.45	6.58–8.30	4.25–9.4	1)
Sulfate ( $SO_4^{2-}$ ) (mg L <sup>-1</sup> )		81–143	13–19	5–100	2)
Ammonia (mg L <sup>-1</sup> )		0.02–0.05	0.001–0.01	<1.57	3)
Dissolved oxygen (mg L <sup>-1</sup> )		5.53–6.10	5.68–6.25	>5	1)
Temperature (°C)		28.42–30.02	28.96–30.07	26–32	1)

1) Courtenay & Williams (2004), 2) Boyd (1988), 3) Jianguang et al (1997), \*) tolerance range, \*\*) optimum range. Table 3 show that the highest survival (73.89%), growth rate (4.40 % day<sup>-1</sup>), albumin content (3.75 g/100 mL), feed efficiency (59.1%), protein retention (24.31%) and energy retention (41.34%) were found on the rainwater medium. The lowest glucose level (26.45 mg/100 mL) was obtained in the rainwater medium.

Table 3 Survival (SR), specific growth rate (SGR), blood albumin level, feed efficiency (FE), protein retention (PR), energy retention (ER), and blood glucose level (BG) of *Channa striata* reared in the different mediums during the study

Parameters	Treatments	Acid sulfate water	Rainwater
SR (%)		54.44±6.21a	73.89±6.12b
SGR (% day <sup>-1</sup> )		2.93±0.40a	4.40±0.96b
Blood albumin (g/100 mL)		3.63±0.65a	3.75±0.66a
FE (%)		28.2±5.41a	59.1±18.76b
PR (%)		5.81±2.16a	24.31±11.12b
ER (%)		10.60±2.95a	41.34±18.28b
BG (mg/100 mL)		29.99±0.58a	26.45±1.13b

Different superscript letters in the same row indicate significant statistical differences (P<0.05).

The high survival and the growth of specimens reared in the rainwater medium related to the water quality conditions of the medium. Water quality parameters (dissolved oxygen, temperature, ammonia, sulfate and pH) of this medium were in the optimum ranges and the tolerance ranges (Table 2). The water temperature in the acid sulfate water medium ranged from 28.42 to 30.02°C, while that in the rainwater medium ranged from 28.96 to 30.07°C, those values did not show any difference to the temperature range required by the species.

Muslim (2007) stated that the temperature range that could be tolerated by the *C. striata* were 25.5-32.7°C. *C. striata* is tolerant to poor water quality conditions. Oxygen is an important factor that determines the life of aquatic organisms. In this study, the dissolved oxygen level in the acid sulfate water medium ranged from 5.53 to 6.10 mg L<sup>-1</sup>, while that of the rainwater medium ranged from 5.68 to 6.25 mg L<sup>-1</sup>. According to Courtenay & Williams (2004), *C. striata* can survive in the low dissolved oxygen level that is less than 5 mg L<sup>-1</sup>.

The dissolved oxygen levels in the treatment mediums during the study were on the optimum range for *C. striata*. This fish is categorized as a fish which has an ability to take oxygen directly from the air. Ammonia (NH<sub>3</sub>) is generated from the reform process of proteins (amino acids) contained in the feed eaten by the fish and converting them into energy.

The metabolic waste is disposed in the form of ammonia (NH<sub>3</sub>) through the gills. In the treatment using acid sulfate water medium, the range of NH<sub>3</sub> contained in the medium was 0.02-0.05 mg L<sup>-1</sup>, while that in the rainwater medium ranged from 0.001 to 0.01 mg L<sup>-1</sup>. These ranges were low and could be tolerated by specimens. At an ammonia concentration of 1.57 mg L<sup>-1</sup>, *C. striata* juvenile can still grow (Jianguang et al 1997). The ranges of sulfate content in the acid sulfate water medium and the rainwater medium were 81-143 mg L<sup>-1</sup> and 13-19 mg L<sup>-1</sup>, respectively. Although the sulfate content in acid sulfate water medium was relatively high, but it was still within the tolerance range for the species (Boyd 1988).

The range of pH during the study in the rainwater medium (6.58-8.30) was still within the tolerance range to promote the growth of *C. striata* juvenile, because this range created an optimum environmental condition to support the physiological activity of the fish. According to Courtenay & Williams (2004), the tolerance range of pH for the life of *C. striata* is at a range of 4.25-9.4.

At this range, the gills ability to bind oxygen becomes more optimal. On the other hand, the acid sulfate water medium showed a pH range below the tolerance range (3.7-5.45), so that the growth of specimens in this medium did not excelled. Although

*C. striata* is sensitive to changes in pH, this species can survive on acidic and alkaline water (Pillay 1995), but the growth will not ensure a maximum level. The low pH and high aluminum concentration in the acid sulfate water may kill the fish or weaken them, so that they become easy to be attacked by diseases and parasites (Singh & Poernomo 1984).

The result of t test presented in Table 3 demonstrated that the survival of *C. striata* juveniles reared in the rainwater medium with a value of 73.89% was significantly different ( $P < 0.05$ ) from those reared in the acid sulfate water medium. It was higher than that on the acid sulfate water medium treatment (54.44%), and it also was higher than the survival of the catfish reared in the freshwater medium as reported in the study of Hollerman & Boyd (1980).

This was caused by the optimum water quality in the rainwater medium that supported the life of *C. striata* compared to acid sulfate water medium, indicated by the ideal water ecosystem both on the feed absorption efficiency and the availability of an optimum amount of dissolved oxygen.

The low survival found in the treatment of acid sulfate water medium was caused by the high energy required by the fish to adapt to environmental stress caused by the low pH. The high energy requirement to survive will stimulate the glucose mobilization to the blood (Costas et al 2008). This was indicated by the high blood glucose level in the treatment of acid sulfate water medium (29.99 mg/100 mL).

The statistical analysis result showed that the treatment of rainwater medium significantly affected ( $P < 0.05$ ) the specific growth rate of *C. striata* juvenile. The specific growth rate data showed that the highest growth of *C. striata* juvenile after 40-day experiment was obtained in the treatment of rainwater medium (4.40%), while the lowest growth was obtained in the treatment of acid sulfate water medium (2.93%).

This showed that the rearing mediums gave a significant effect on the fish growth. The specimens reared in the rainwater medium were better in utilizing energy obtained from the feed, so the nutrients contained in the feed could be metabolized efficiently. A better growth of specimens reared in the rainwater medium was reflected by a high protein retention value (24.31%).

Ballestrazzi et al (1994) stated that the protein retention was a parameter to indicate the contribution of protein consumed from the diet on the body protein accretion. Protein retention needs to be specifically noted to see the contribution of protein consumed from the diet to the increase of the fish weight or height (Watanabe 1988).

The protein retention value also shows the quality of protein contained in the diet, the higher the protein retention value, the better the feed quality (Halver 1989). It was clear that the rainwater medium contributed better to the feed energy utilization, as more proteins were stored and were decomposed only in a little amount or those were utilized as energy to maintain homeostasis.

Lipid is usually stored as energy reserves for long-term energy needs during the full activity period or during the period without food and energy. This phenomenon could be seen from the energy retention of *C. striata* juvenile. The fish reared in the rainwater medium had an energy retention value of 41.34%, while the result obtained in the treatment of acid sulfate water medium was 10.60%.

The low energy retention in the treatment of acid sulfate water medium possibly happened because the energy produced secreted more by the body for metabolism, reproductive activity, biosynthesis and it was also lost as heat. Moreover, Jobling et al (1991) stated that the high energy for the activity of the organism will reduce the energy budget for growth.

According to Nelson & Chabot (2011), the energy in the feed is physiologically used for the body treatment and metabolism, if there is a rest of it, it will be deposited as the body tissues in the process of growth and reproductive tissue synthesis. The results showed that the higher value of feed efficiency is on the treatment of the rainwater medium (59.1%) compared to the treatment of acid sulfate water medium (28.2%). The high feed efficiency in the treatment of rainwater medium indicated a better feed utilization.

The efficiency of feed utilization will run optimally, if the environmental conditions were in normal conditions. According to Brett (1971), the amount of feed consumed by the fish per day is one of the factors that influence the potential of the fish to grow maximally and the daily feed intake is closely related to the capacity and the emptying of the stomach.

The daily feeding rate, feeding frequency and type of feed used affect the digestibility of the fish, which plays an important role in lowering feed conversion ratio or increasing feed efficiency by efficient utilization of the feed (Chiu et al 1987; De Silva & Davy 1992). The results of the present study demonstrated that the blood albumin level of the fish reared in the rainwater medium did not show any difference with those reared in the acid sulfate water medium. *C. striata* is one of the species which has a high level of blood albumin (Mustafa et al 2012). Albumin is a kind of globular protein which soluble in water, salt and acid solvents (Masuelli 2013). The previous studies reported that fish physiological status, age, season and habitats

influenced on serum protein properties, especially albumin (Kovyrshina & Rudneva 2012). Several researchers have shown that albumin concentration strongly depends on diet composition (Chukwuma et al 2010; MacQueen et al 2011). Moreover, fish physiological status that affect albumin concentration strong correlated with seasonal fluctuations depending on water temperature, oxygen level, food composition, anthropogenic impact and algae bloom (Kamal & Omar 2011). In this study, diet composition, water temperature, and oxygen level were in the same range, so that the blood albumin levels of *C. striata* specimens reared in the different mediums tested in this study did not show any differences. Conclusions. The use of the rainwater as a rearing medium for *C. striata* juveniles gave higher values concerning all indices monitored (SR, SGR, FE, PR, ER, BG), against acid sulfate water medium.

References Ballestrazzi R., Lannari D., D'Agaro E., Mion A., 1994 The effect of dietary protein level and source on growth, body composition, total ammonia and reactive phosphate excretion of growing sea bass (*Dicentrarchus labrax*). *Aquaculture* 127:197-206.

Boyd C. E., 1988 Water quality in warmwater fish ponds. Agricultural Experiment Station, Auburn University, Alabama, 359 pp.

Brett J. R., 1971 Satiation time, appetite, and maximum food intake of sockeye salmon (*Oncorhynchus nerka*). *Journal of the Fisheries Research Board of Canada* 28(3):409-415. Chandra S., Banerjee T. K., 2004 Histopathological analysis of the respiratory organs of *Channa striata* subjected to air exposure. *Veterinarski Arhiv* 74(1):37-52.

Chiu Y. N., Sumagaysay N. S., Sastrillo M. A. S., 1987 Effect of feeding frequency and feeding rate on the growth and feed efficiency of milkfish, *Chanos chanos* Forsskal, juveniles. *Asian Fisheries Science* 1(1):27-31.

Chukwuma E. R., Obioma N., Christopher O. I., 2010 The phytochemical composition and some biochemical effects of Nigerian tigernut (*Cyperus esculentus* L.) tuber. *Pakistan Journal of Nutrition* 9(7):709-715.

Costas B., Aragão C., Mancera J. M., Dinis M. T., Conceição L. E. C., 2008 High stocking density induces crowding stress and affects amino acid metabolism in Senegalese sole *Solea senegalensis* (Kaup 1858) juveniles. *Aquaculture Research* 39(1):1-9.

Courtenay W. R., Williams J. D., 2004 Snakeheads (Pisces, Channidae): A biological synopsis and risk assessment. U.S. Geological Survey, Reston, 143 pp.

De Silva S. S., Davy F. B., 1992 Fish nutrition research for semi-intensive culture systems in Asia. *Asian Fisheries Science* 5:129-144.

El-Sayed A.-F. M., El-Ghobashy A., Al-Amoudi M., 1996 Effects of pond depth and water temperature on the growth, mortality and body composition of Nile tilapia, *Oreochromis niloticus* (L.). *Aquaculture Research* 27(9):681-687.

Halver J. E., 1989 Fish nutrition, 2nd edition. Academic Press, New York, 798 pp.

Hidayatullah S., Muslim, Taqwa F. H., 2015 [Rearing of snakehead larvae (*Channa*

striata) in plastic lined pond with different stocking density]. *Jurnal Perikanan dan Kelautan* 20(1):61-70. [In Indonesian].

Hollerman W. D., Boyd C. E., 1980 Nightly aeration trincreeye production of channel catfish. *Transaction of the American Fisheries Society* 109:440-452.

Infusino I., Panteghini M., 2013 Serum albumin: accuracy and clinical use. *Clinica Chimica Acta* 419:15-18.

Jais A. M. M., Dambisya Y. M., Lee T. L., 1997 Antinociceptive activity of *Channa striatus* (Haruan) extracts in mice. *Journal of Ethnopharmacology* 57(2):125-130.

Jianguang Q., Fast A. W., Kai A. T., 1997 Tolerance of snakehead *Channa striatus* to ammonia at different pH. *Journal of the World Aquaculture Society* 28(1):87-90.

Jobling M., Knudsen R., Pedersen P. S., Dos Santos J., 1991 Effects of dietary composition and energy content on the nutritional energetics of cod, *Gadus morhua*. *Aquaculture* 92:243-257. Kamal S. M.,

Omar W. A., 2011 Effect of different stocking densities on hematological and biochemical parameters of silver carp, *Hypophthalmichthys molitrix* fingerlings. *Life Science Journal* 8(4):580-586.

Kang'ombe J., Brown J. A., 2008 Effect of salinity on growth, feed utilization, and survival of *Tilapia rendalli* under laboratory conditions. *Journal of Applied Aquaculture* 20(4):256-271.

Kovyrshina T. B., Rudneva I. I., 2012 Comparative study of serum albumin levels in round goby *Neogobius melanostomus* from Black Sea and Azov Sea. *International Journal of Advanced Biological Research* 2(2):203-208.

MacQueen H. A., Wassif W. S., Walker I., Sadler D. A., Evans K., 2011 Age-related biomarkers can be modulated by diet in the rat. *Food and Nutrition Sciences* 2(8):884-890.

Masuelli M. A., 2013 Study of bovine serum albumin solubility in aqueous solutions by intrinsic viscosity measurements. *Advances in Physical Chemistry*, Article ID 360239, 8 pp, <http://dx.doi.org/10.1155/2013/360239>.

Mollah M. F. A., Mamun M. S. A., Sarowar M. N., Roy A., 2009 Effects of stocking density on the growth and breeding performance of broodfish and larval growth and survival of shol, *Channa striatus* (Bloch). *Journal of the Bangladesh Agricultural University* 7(2):427-432.

Muslim, 2007 [Potential, opportunity and challenge of the culture of the snakehead fish (*Channa striata*) in South Sumatera Province]. *Proceeding IV Indonesian Public Water Forum IV, Palembang*, pp. 7-11. [In Indonesian].

Mustafa A., Widodo M. A., Kristianto Y., 2012. Albumin and zinc content of snakehead fish (*Channa striata*) extract and its role in health. *IEESE International Journal of Science and Technology* 1(2):1-8.

Nelson J. A., Chabot D., 2011 General energy metabolism. In: *Encyclopedia of fish physiology: from genome to environment*, volume 3. Farrell A. P. (ed), pp. 1566-1572, Academic Press, San Diego. Ng P. K. L., Lim K. K. P., 1990 Snakeheads (Pisces: Channidae): Natural history, biology and economic importance. *Essays in Zoology*,

Papers Commemorating the 40th Anniversary of the Department of Zoology, National University of Singapore, Singapore, pp. 127-152.

Pillay T. V. R., 1995 Aquaculture principles and practices. Fishing News Books, Cambridge, 575 pp. Purnamawati, 2017 [The growth performance of snakehead *Channa striata* Bloch. on tidal land through water quality engineering]. Bogor Agricultural University, Bogor, Indonesia, 78 pp. [In Indonesian].

Qin J., Fast A. W., DeAnda D., Weidenbach R. P., 1997 Growth and survival of larval snakehead (*Channa striatus*) fed different diets. *Aquaculture* 148(2-3):105-113.

Singh V. P., Poernomo A. T., 1984 Acid sulfate soils and their management for brackishwater fishponds. *Advances in Milkfish Biology and Culture, Proceedings of the Second International Milkfish Aquaculture Conference, Iloilo City*, pp. 121-132.

Song L. M., Munian K., Rashid Z. A., Bhassu S., 2013 Characterisation of Asian snakehead murrel *Channa striata* (Channidae) in Malaysia: an insight into molecular data and morphological approach. *The Scientific World Journal*, Article ID 917506, 16 pages <http://dx.doi.org/10.1155/2013/917506>.

Takeuchi T., 1988 Laboratory work-chemical evaluation of dietary nutrient. In: *Fish nutrition and mariculture*. Watanabe T. (ed), pp. 179-233, Kanagawa International Fisheries Training Centre, Japan International Cooperation Agency (JICA), Tokyo.

Vivekanandan E., 1977 Surfacing activity and food utilization in the obligatory air-breathing fish *Ophiocephalus striatus* as a function of body weight. *Hydrobiologia* 55(2):99-112.

Watanabe T., 1988 Fish nutrition and mariculture. In: *Live feeds, JICA textbook, the general aquaculture course*. Kanagawa International Fisheries Training Centre, Japan International Cooperation Agency, Tokyo, 233 pp.

Weatherley A. H., Gill H. S., 1989 *The biology of fish growth*. Academic Press, London, 443 pp.

Wedemeyer G. A., Yasutake W. T., 1977 *Clinical methods for the assessment of the effects of environmental stress on fish health*. U.S. Fish and Wildlife Service, Washington D.C., 18 pp.

\*\*\* APHA (American Public Health Association), 1989 *Standard methods for the examination of water and wastewater*. 17th edition. American Public Health Association, Washington D.C., 1268 pp. Received: 13 February 2017. Accepted: 19 March 2017. Published online: 23 March 2017. Authors:

Purnamawati, Bogor Agricultural University, Graduate School, Study Program of Aquaculture, Indonesia, West Java, Bogor 16680, Dramaga Campus; Pontianak State Polytechnic University, Department of Marine Science and Fisheries, Study Program of Aquaculture, Indonesia, West Borneo, Pontianak 78124, Jalan Ahmad Yani, e-mail: [purnamawati\\_z@yahoo.com](mailto:purnamawati_z@yahoo.com) Daniel Djokosetiyanto, Bogor Agricultural University, Faculty of Fisheries and Marine Science, Department of Aquaculture, Indonesia, West Java, Bogor 16680, Dramaga Campus, e-mail: [djokosetiyanto@yahoo.co.id](mailto:djokosetiyanto@yahoo.co.id) Kukuh Nirmala, Bogor Agricultural University, Faculty of Fisheries and Marine Science, Department of Aquaculture, Indonesia, West Java, Bogor 16680, Dramaga Campus,

e-mail: kukuhnirmala@yahoo.com Enang Harris, Bogor Agricultural University, Faculty of Fisheries and Marine Science, Department of Aquaculture, Indonesia, West Java, Bogor 16680, Dramaga Campus, e-mail: enangharris@yahoo.com Ridwan Affandi, Bogor Agricultural University, Faculty of Fisheries and Marine Science, Department of Aquatic Resources Management, Indonesia, West Java, Bogor 16680, Dramaga Campus, e-mail: affandi\_ridwan@yahoo.com This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited. How to cite this article: Purnamawati, Djokosetiyanto D., Nirmala K., Harris E., Affandi R., 2017 Survival and growth of striped snakehead fish (*Channa striata* Bloch.) juvenile reared in acid sulfate water and rainwater medium. AACL Bioflux 10(2):265-273.

#### INTERNET SOURCES:

-----  
<http://www.bioflux.com.ro/docs/2017.265-273.pdf>

<http://reefkeeping.com/issues/2005-10/rhf/>

<https://www.sciencedirect.com/science/article/pii/S0044848608003062>